

# STUDY OF NUTRITIONAL LIMITATIONS OF WHEAT BRAN (WB) BASED CULTURE MEDIUM FOR SCALING UP *BTK* BASED BIOPESTICIDE PRODUCTIONS

---

Rita BARSSOUM





1

---

# Scientific context

# 1.1. IPM-4-Citrus (MSCA RISE, No. 734921, 2017-2023)

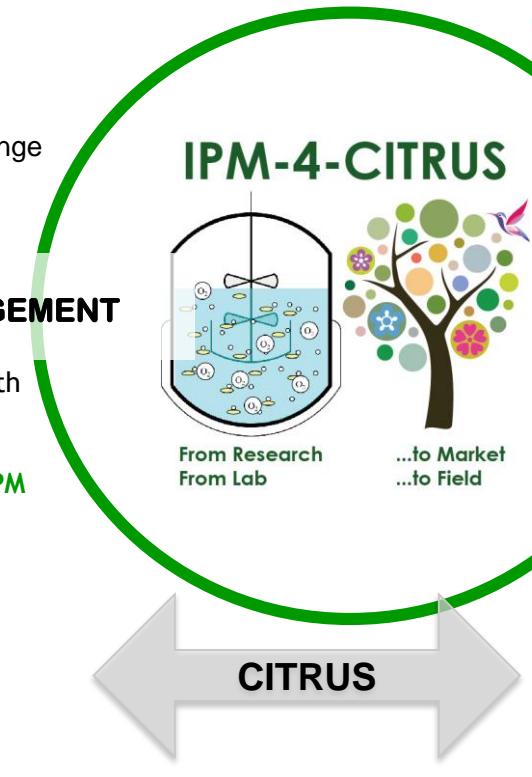
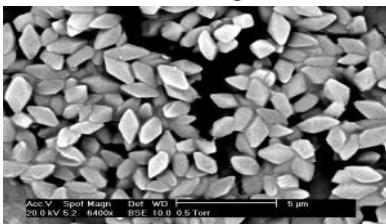
HORIZON 2020 FUNDED

Marie Skłodowska Curie Action

Research & Innovation Staff Exchange

## INTEGRATED PEST MANAGEMENT

- ✓ Understanding & sensitising stakeholders about the health risks related to citrus pests
- ✓ Developing an alternative IPM approach
- ✓ based on biological control



11 PARTNERS

6 COUNTRIES

4 YEARS DURATION



LISBP  
Institut des Systèmes Biologiques  
et des Procédés



TARGETED PEST:  
*insect larvae : Phyllocnistis citrella & Prays citri*



This project has received funding from the European Union's Horizon 2020 Research and Innovation program under Grant Agreement No 734921.

# 1.2. *Bacillus thuringiensis kurstaki* (Btk)

*Bacillus thuringiensis kurstaki*  
(LIP, HD1, BLB1)

Bacilli

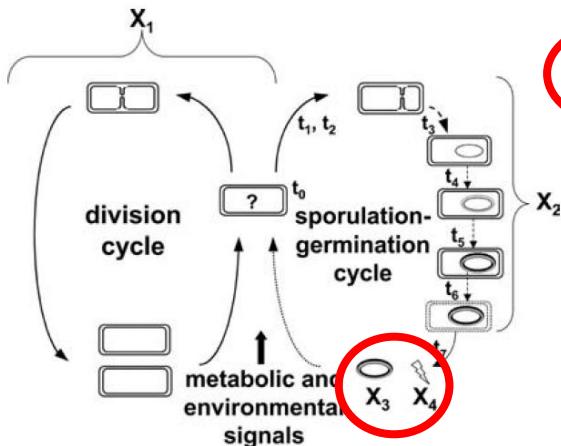
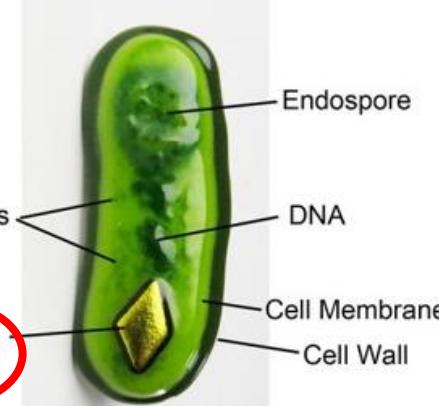
Gram +

Facultative aeroby

Sporulant

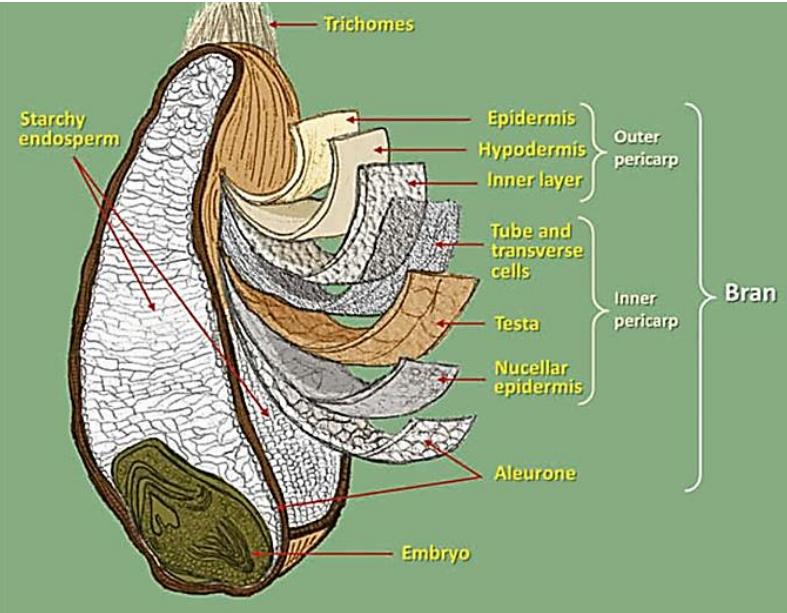
Endotoxin  
producer

*Bacillus thuringiensis* (Bt) life cycle and endotoxins production (<https://www.bpprc.org/>)



(M. Sarrafzadeh et al 2005)

# 1.3. Wheat bran (WB) structure & chemical composition



Wheat Bran Kernel structure  
(Balandran et al, 2015)

Mean composition of Wheat Bran (WB)  
(Hell et al., 2016, Sapirstein et al, 2016, Stoffel et al., 2019)

Component		Content in WB (%w/w)
Polysaccharides	Reserve sugars <ul style="list-style-type: none"><li>- Oligosaccharides</li><li>- Starch</li></ul>	15-45 3,7 13-40
	Fibres (structure) <ul style="list-style-type: none"><li>- Cellulose</li><li>- Hemicellulose</li><li>- Lignin</li></ul>	35,7-62,3 6,5-11 20,8-33 9,8-16
Proteins		13,2-21,1
Ashes/Minerals		2,2-8
Water		9-12

## 1.4. Background

WB: good medium  
for Bt crystals  
production (Devi et  
al, 2005)

Effective low cost Btk crystal  
production in cheap WB  
based medium compared to  
semi synthetic medium  
(Mounsef et al, 2014)

Sieving of WB into 4 classes  
: class 1 ( $>850\text{ }\mu\text{m}$ ), class 2  
( $500\text{-}850\text{ }\mu\text{m}$ ), class 3 (250-  
 $500\text{ }\mu\text{m}$ ) & class 4 ( $<250\text{ }\mu\text{m}$ ).

Optimal Btk growth &  
crystal production in class 2,  
([WB]=73.6 g/L), 248 rpm  
(Abboud et al, 2017)

## 1.4. Main Scientific Questions

1. Does the substrate elemental composition differ between one class and another?
2. Do the mass balance and the elemental composition inform us about the progress of the culture, the limiting nutrients and fermentable fraction?
3. In terms of scale-up approach, are the limiting components in bioreactor culture, the same as in flask culture?
4. Can the elemental composition help us interpreting the 3 phases of culture & biokinetics in bioreactor?

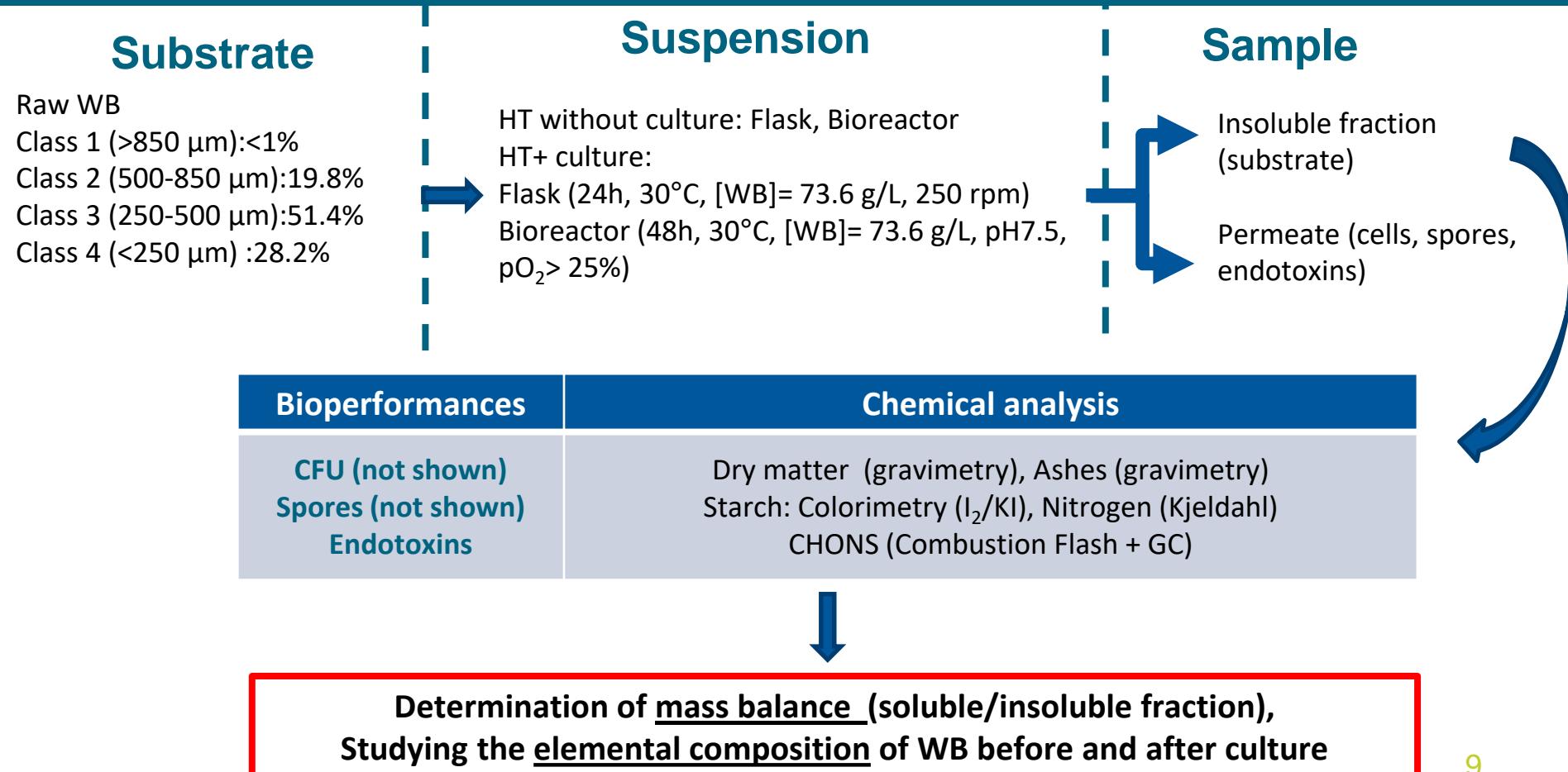


2

---

## Materials & Methods

## 2. Materials & Methods





3

---

## Results & Discussion

### 3.1. Endotoxins production in WB vs Semi-synthetic medium (SSM)

*Btk* endotoxins yield in mg/mL

Btk Strains	Culture in bioreactor		Culture in flasks		
	SSM Sarrafzadeh et al, 2005	WB Raw Rahbani et al, 2015	WB Class 2	WB Class 3	WB Class 4
HD1	0,7	0,43	/	/	/
BLB1	0,73	0,63	/	/	/
Lip	0,19	0,43	0,549 ±0,117	0,547 ±0,071	0,432 ±0,062

- *Btk* Lip: **Higher yield** in WB vs SSM
- *Btk* Lip bioperformances ↑ on class 2 & 3

## 3.2. Raw and sieved WB composition

			Mass	Water content	Starch	Protein Kjeldahl	Elemental Composition [%w/w]					
Class	Size ( $\mu\text{m}$ )	D [4,3] ( $\mu\text{m}$ )	[%w/w]	[g water /gdm]	[g/gdm]	[g eq.protein /gdm]	C	H	O	N	S	Ash
Raw		598,5	100	0,119 $\pm 0,07$	0,207 $\pm 0,008$	0,138 $\pm 0,012$	44,79 $\pm 0,44$	6,74 $\pm 0,20$	36,53 $\pm 0,37$	2,50 $\pm 0,24$	0,00	4,43 $\pm 0,12$
2	500-850	865,4	19,80	0,116 $\pm 0,04$	0,173 $\pm 0,35$	0,145 $\pm 0,012$	44,21 $\pm 0,19$	6,50 $\pm 0,12$	35,62 $\pm 0,45$	2,60 $\pm 0,23$	0,00	3,70 $\pm 0,12$
3	250-500	531,1	51,41	0,116 $\pm 0,04$	0,144 $\pm 1,35$	0,127 $\pm 0,014$	45,34 $\pm 0,08$	6,46 $\pm 0,04$	37,89 $\pm 0,44$	2,48 $\pm 0,04$	0,00	4,28 $\pm 0,12$
4	<250	275,3	28,20	0,116 $\pm 0,04$	0,347 $\pm 0,47$	0,155 $\pm 0,016$	42,40 $\pm 0,04$	6,61 $\pm 0,12$	34,60 $\pm 0,37$	2,61 $\pm 0,24$	0,00	3,23 $\pm 0,12$

- WB granulometry → Ø effect on elemental composition
- WB molar formula (mol/C\_mol):  $\text{CH}_{1,74}\text{O}_{0,53}\text{N}_{0,04}$
- WB massic formula (g/g\_C):  $\text{CH}_{0,14}\text{O}_{0,71}\text{N}_{0,05}\text{Ash}_{0,1}$
- Class 4: highest starch [ ]

### 3.3. *Btk* elemental composition

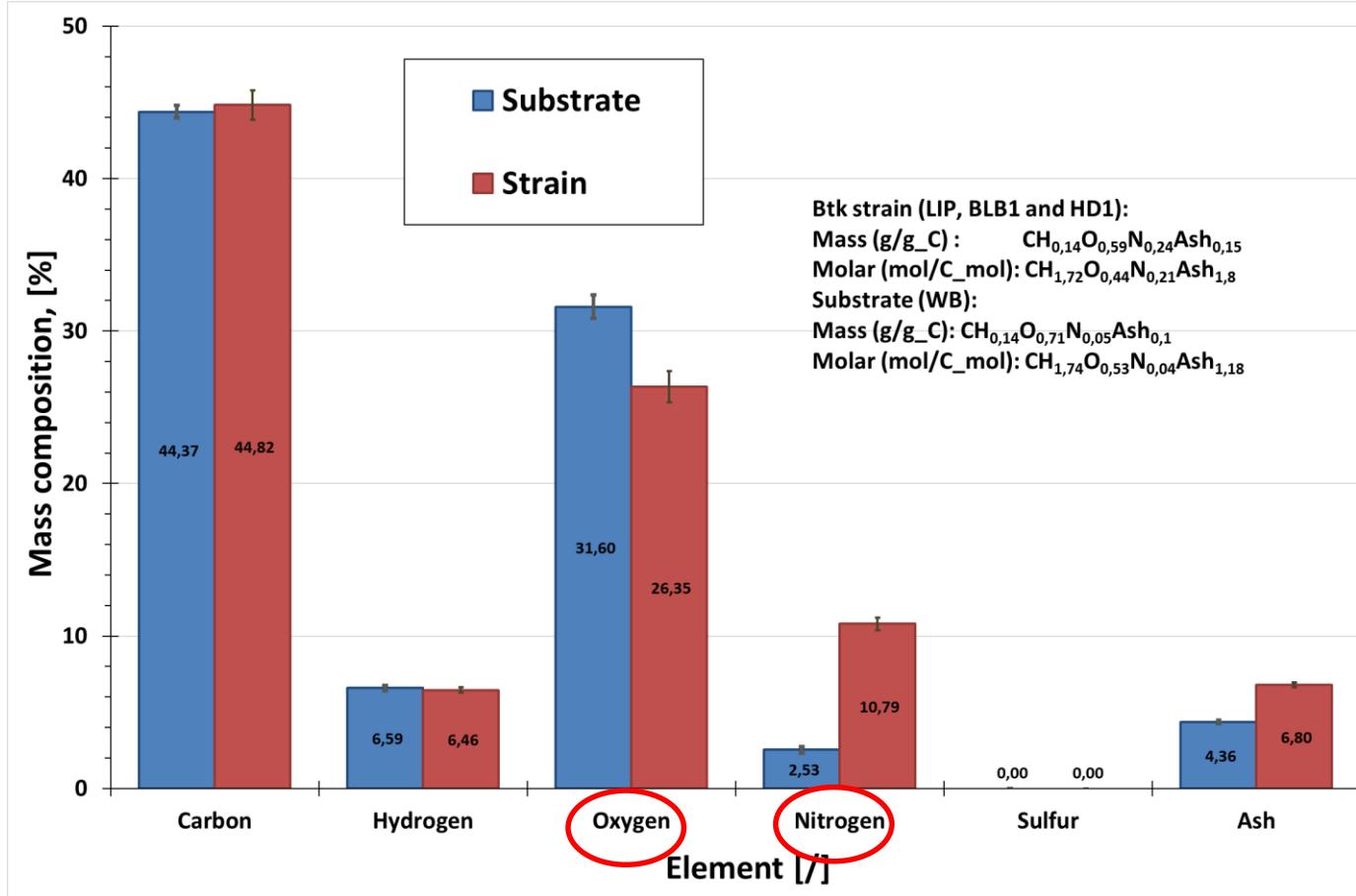
Btk Strains	C (%w/w)	H (%w/w)	O (%w/w)	N (%w/w)	S (%w/w)	Ashes (%w/w)
Lip	43,37	6,22	24,84	10,48	0, 00	7
BLB1	45,7 ±0,067	6,61 ±0,09	26,89 ±0,87	10,51 ±0,011	0, 00	6,7
HD1	44,65 ±0,3	6,42 ±0,072	26,56 ±0,68	11,23 ±0,068	0, 00	NA
Mean value	44,82 ±0,98	6,46 ±0,17	26,35 ±1	10,79 ±0,4	0, 00	6,8 ±0,17

- *Btk* strains: identical elemental composition
- **Molar formula of the strain (mol/C\_mol):**  $\text{CH}_{1,72}\text{O}_{0,44}\text{N}_{0,21}$
- **Massic formula of the strain (g/g\_C):**  $\text{CH}_{0,14}\text{O}_{0,59}\text{N}_{0,24}\text{Ash}_{0,15}$

Strain	Elements (%m/m)					Chemical formula	Molar Mass
	Ashes	C	H	O	N		
<i>Bt spp. kurstaki (MSc)</i>	6,80	44,82	6,46	26,35	10,79	$\text{CH}_{1,72}\text{O}_{0,44}\text{N}_{0,21}$	25,52
<i>Lactobacillus helveticus</i>	9,03	47,54	6,25	24,39	12,79	$\text{CH}_{1,58}\text{O}_{0,39}\text{N}_{0,23}$	25,26
<i>Flavobacterium Dehydrogenans</i>	13,5	45,16	6,15	24,29	10,87	$\text{CH}_{1,63}\text{O}_{0,40}\text{N}_{0,21}$	26,60
<i>Escherichia coli</i>	11,3	47,83	6,95	21,65	12,3	$\text{CH}_{1,74}\text{O}_{0,34}\text{N}_{0,22}$	25,11
<i>Bacillus cereus</i>	9,98	46,05	5,73	26,26	11,98	$\text{CH}_{1,49}\text{O}_{0,43}\text{N}_{0,22}$	26,08

➤ *Btk*: elemental composition close to *Bacillus cereus* (Popovic et al, 2019)

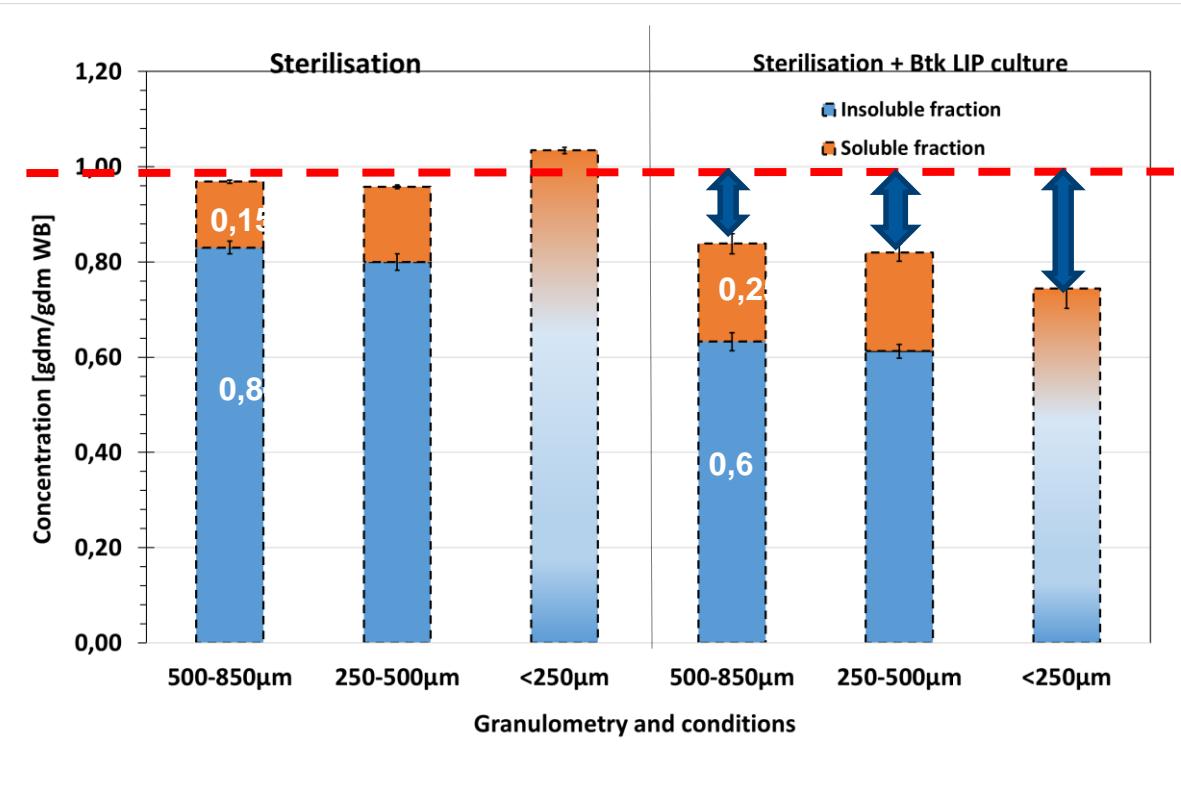
### 3.4. Btk vs WB, Elemental composition



- Strains are more rich in Nitrogen and less in Oxygen than WB
- WB is the **only source of Nitrogen** during Btk culture

### 3.5.What is the fermentable fraction over classes?

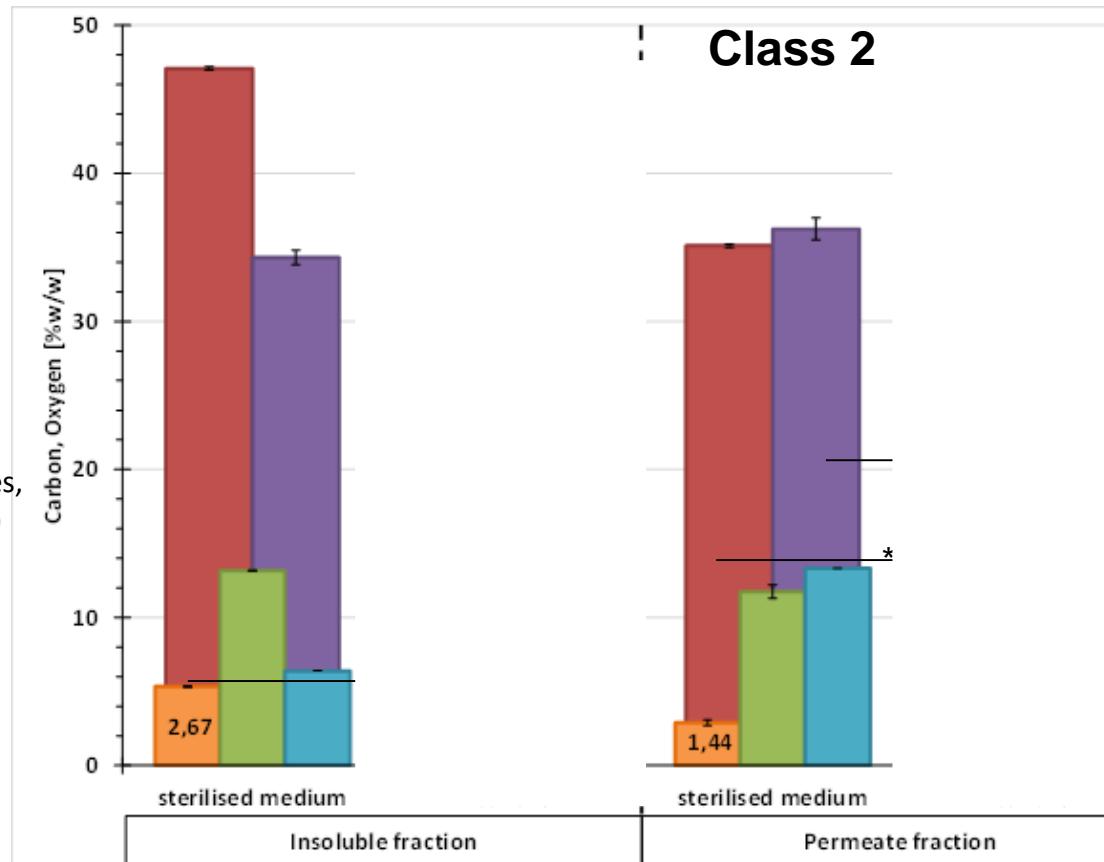
Substrat + Oxygène + Azote + Oligo + ... → Biomasse + CO<sub>2</sub> + produits (Spore, endotoxine) + eau + énergie  
1/6 C<sub>6</sub>H<sub>12</sub>O<sub>6</sub> + α O<sub>2</sub> + β NH<sub>3</sub> + Sels +... → Y<sub>sx</sub> X (CH<sub>1,8</sub>O<sub>0,3</sub>N<sub>0,2</sub>) + δ CO<sub>2</sub> + λ H<sub>2</sub>O + ε Spores + ξ Endotoxine + Energie...



- ↗ fermentable fraction when granulometry ↘
- If Y<sub>x/s</sub> = 0,5 fermentable fraction ~ Starch
  - ➔ Class 2 : 32,3%
  - ➔ Class 3 : 36,1%
  - ➔ Class 4 : 51,1%
- Partial consumption of lignocellulosic fraction ([Starch]= 0,173 gdm/gdmWB)

### 3.6. How does the medium elemental composition evolve?

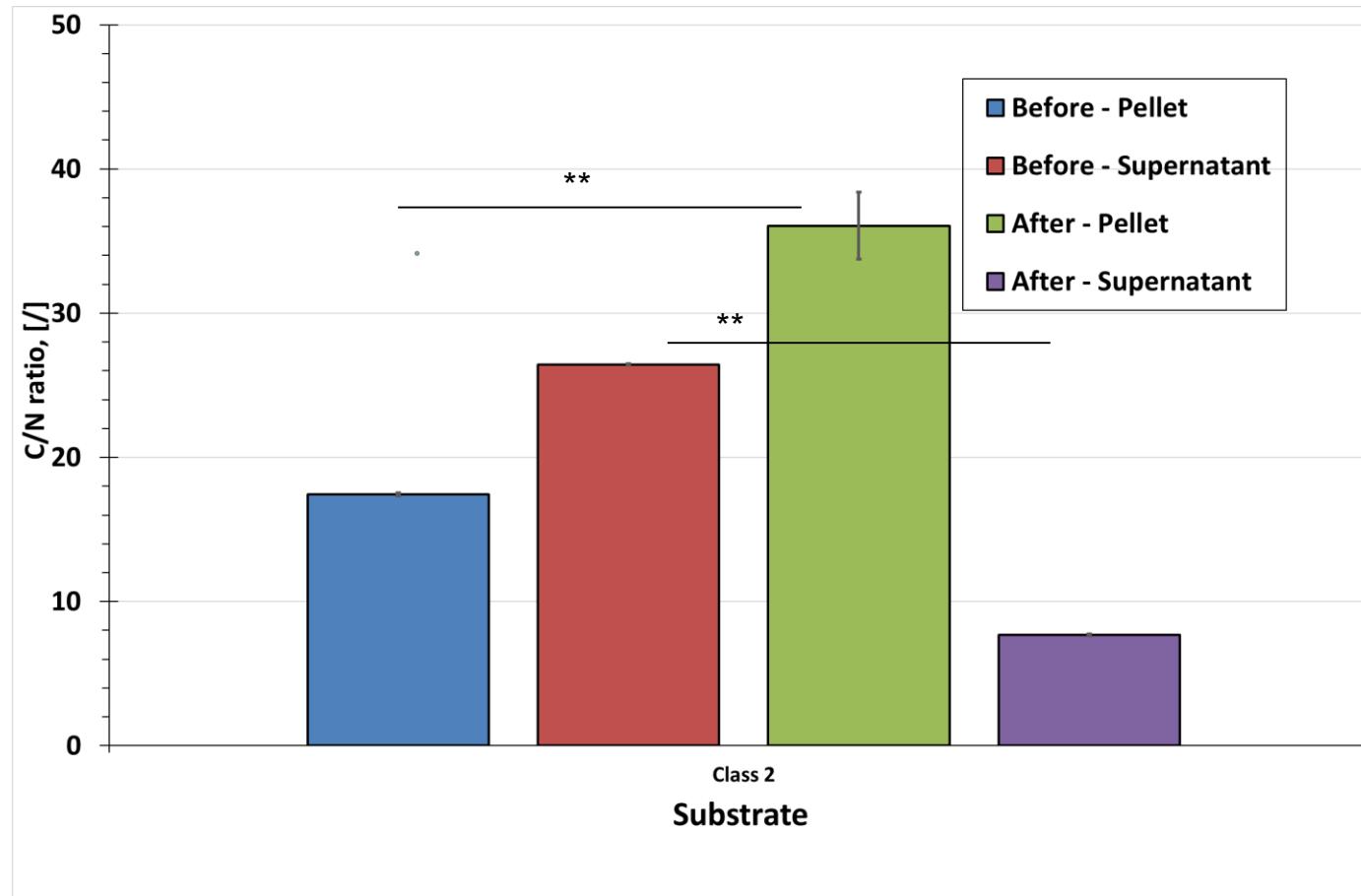
Insoluble fraction (substrate)  
Permeate (cells, spores, endotoxins)



Class 2

- Permeate: Biomass production
- Insoluble fraction: substrate proteins consumption
- N: limiting nutrient
- N disponibile ~ 48% N accessible
- Class 3: same behaviour
- Class 4: No separation pellet/supernatant

### 3.7. C/N ratio: indicator of the process evolution in flask?

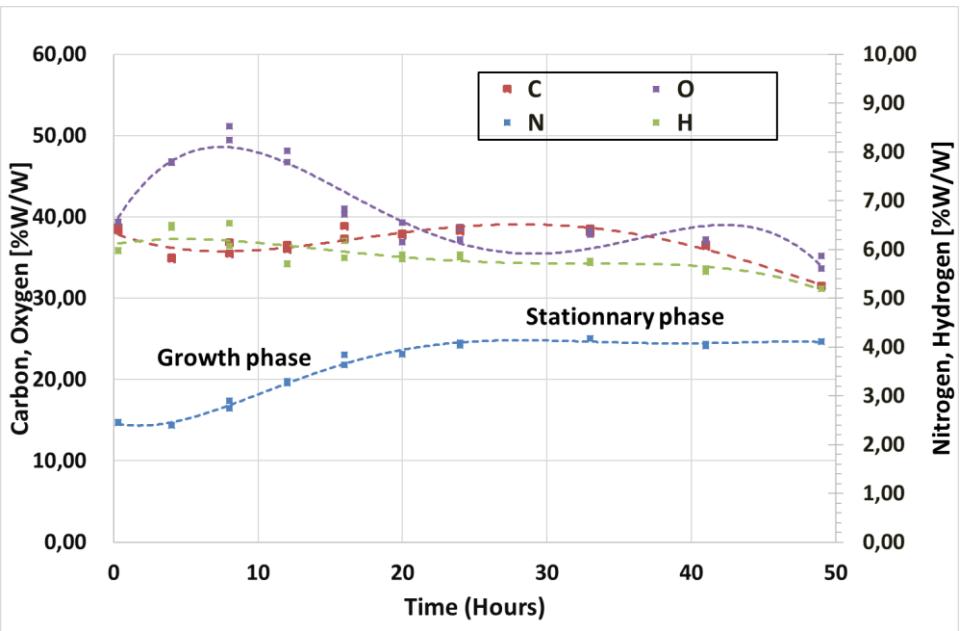


- C/N is a reliable indicator to interpret the process evolution & phases of culture

### 3.8. Monitoring of culture biokinetics through elemental composition analysis in bioreactor

#### Permeate

#### Insoluble fraction



- Same behaviour between flask and bioreactor.
- Biokinetics: 0-12h: growth phase, 20- 48h: stationary phase followed by sporulation phase

# 4

---

## Conclusion & Perspectives

## 4.1. Highlights & perspectives

- WB elemental composition: no evolution between classes
- Fermentable fraction: starch & lignocellulosic fraction
- N: culture limiting nutrient
- Residual N: Physically non accessible
- C/N ratio: reliable indicator of culture phases & bioperformances (flask & bioreactor scale)

Determination of chemical limitations → optimisation of biopesticide production in a low cost WB medium at large scale

## 4.1. Highlights & perspectives

### Perspectives:

- Study WB physical characteristics (morpho-granulometry, rheometry, settling speed)
- Define an appropriate mathematical model in order to better describe bioperfmances & optimise endotoxin production

# Special thanks to:



## UR- EGP, Saint-Joseph University of Beirut,

Prof Mireille KALLASSY AWAD  
Dr. Nancy FAYAD  
Ms. Joanna ABOUD  
Mr. Rayane NASSERDINNE  
Ms. Gabrielle EL KASSIS  
Ms. Jihane SAAD



## TBI- INSA Toulouse



Dr. Luc FILLAUDEAU  
Dr. César Arturo ACEVES-LARA



## Toulouse White Biotechnology Institute

Dr. Julien CESCUT,  
Mrs. Stéphanie DUPOIRON



## Société MEDIS, Nabeul, Tunisia

Ms. Meriem EL GHOUL

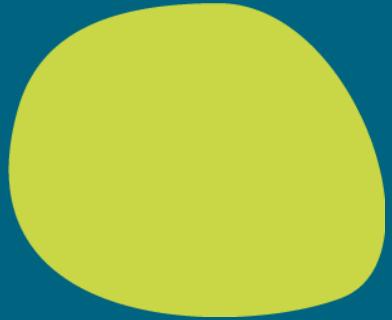


## The organizing committee of BACT2022

Michel GOHAR, Véronique BROUSSOLLE, Leyla SLAMTI, Jean-Nicolas TOURNIER



This project has received funding from the European Union's Horizon 2020 Research and Innovation program under Grant Agreement No 734921.



---

Thank you for your attention

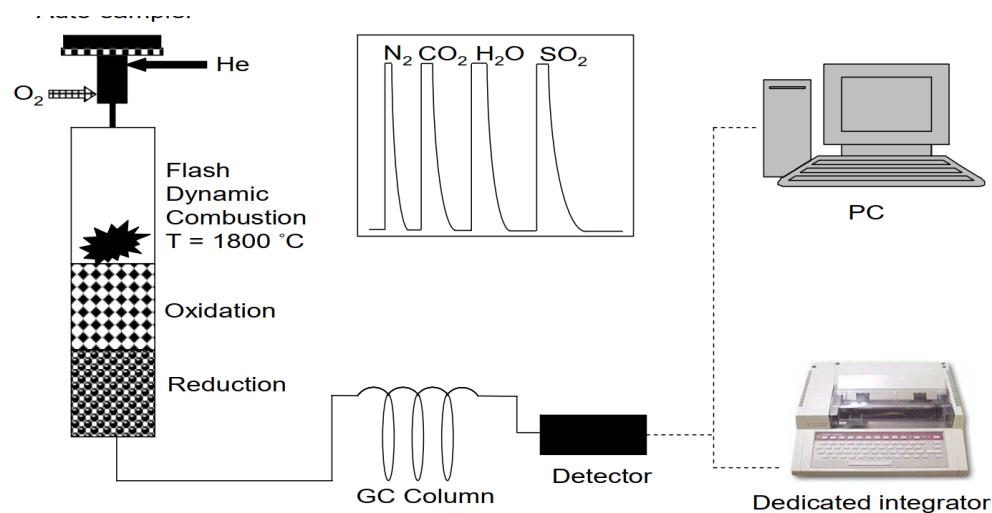


5

---

## Back-up

## 5. CHONS analysis method

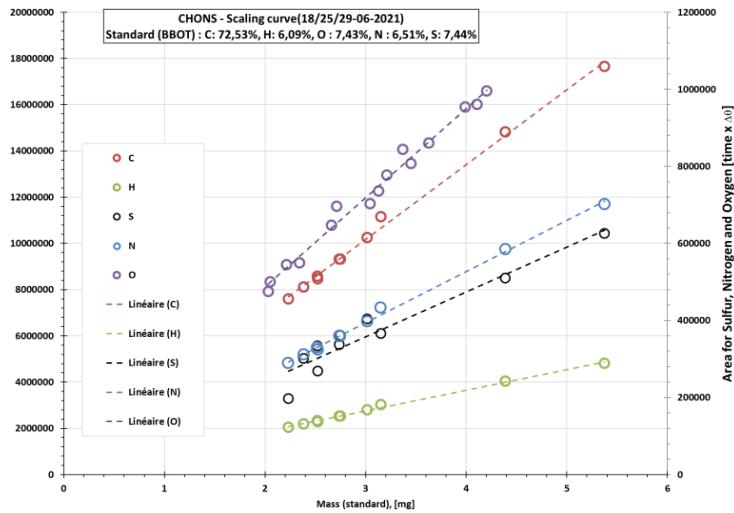


Flash smart 2000, thermofischer scientific

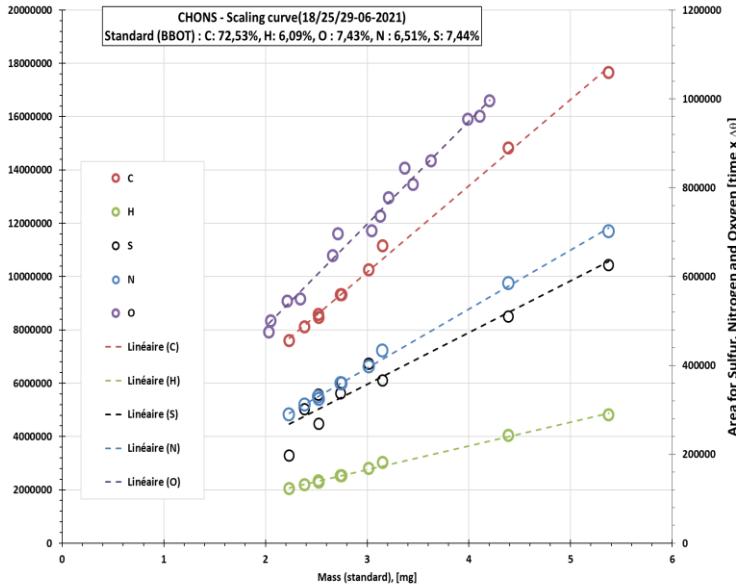
CHONS analysis mechanism

# 5. CHONS calibration curves

Area for Carbon and Hydrogen [time x Δθ]



Area for Carbon and Hydrogen [time x Δθ]



## 8. DM and ashes analysis method



# 6

---

## References

## 6. References

- Balandran et al , Wheat Bran Proteins: A Review of Their Uses and Potential, *Food Reviews International*, 31:279–293, <https://doi.org/10.1080/87559129.2015.1015>
- **Sapirstein, H.D., 2016. Bioactive Compounds in Wheat Bran, in: Wrigley, C., Corke, H., Seetharaman, K., Faubion, J. (Eds.), Encyclopedia of Food Grains (Second Edition). Academic Press, Oxford, pp. 268–276.** <https://doi.org/10.1016/B978-0-12-394437-5.00109-1>
- Palma, L., Muñoz, D., Berry, C., Murillo, J., Caballero, P., 2014. *Bacillus thuringiensis* Toxins: An Overview of Their Biocidal Activity. *Toxins* 6, 3296–3325. <https://doi.org/10.3390/toxins6123296>
- Popovic, M., 2019. Thermodynamic properties of microorganisms: determination and analysis of enthalpy, entropy, and Gibbs free energy of biomass, cells and colonies of 32 microorganism species. *Heliyon* 5, e01950. <https://doi.org/10.1016/j.heliyon.2019.e01950>
- Hell, J., Prückler, M., Danner, L., Henniges, U., Apprich, S., Rosenau, T., Kneifel, W., Böhmdorfer, S., 2016. A comparison between near-infrared (NIR) and mid-infrared (ATR-FTIR) spectroscopy for the multivariate determination of compositional properties in wheat bran samples. *Food Control* 60, 365–369. <https://doi.org/10.1016/j.foodcont.2015.08.003>
- Hemery, Y., Rouau, X., Dragan, C., Bilici, M., Beleca, R., Dascalescu, L., 2009. Electrostatic properties of wheat bran and its constitutive layers: Influence of particle size, composition, and moisture content. *J. Food Eng.* 93, 114–124. <https://doi.org/10.1016/j.jfoodeng.2009.01.003>
- **Sapirstein, H.D., 2016. Bioactive Compounds in Wheat Bran, in: Wrigley, C., Corke, H., Seetharaman, K., Faubion, J. (Eds.), Encyclopedia of Food Grains (Second Edition). Academic Press, Oxford, pp. 268–276.** <https://doi.org/10.1016/B978-0-12-394437-5.00109-1>
- Sarrafzadeh, M.H., Navarro, J.M., 2006. The effect of oxygen on the sporulation, δ-endotoxin synthesis and toxicity of *Bacillus thuringiensis* H14. *World J. Microbiol. Biotechnol.* 22, 305–310. <https://doi.org/10.1007/s11274-005-9037-9>